

***Nigella sativa* Oil Improves Follicular Reserve in Cyclophosphamide-Induced Ovarian Toxicity: A Histomorphological and Hormonal Assessment in Female Rats**

Murtala Muhammad Jibril¹, Mohammed Bello Mohammed², Usman Adamu Garkuwa³, Ede Samuel Otokpa⁴, Ali Ishaq Shugaba⁵, Ekwere O. Ekwere⁶

¹Department of Human Anatomy, Faculty of Basic Medical Sciences
Saadu Zungur University, Bauchi State Nigeria
Email: mmjibril [AT] basug.edu.ng

²Department of Human Anatomy, Faculty of Basic Medical Sciences
University of Jos, Plateau State Nigeria
Email: mohdbello2015 [AT] gmail.com

³Department of Human Physiology, Faculty of Basic Medical Sciences
Saadu Zungur University, Bauchi State, Nigeria
Email: garkuwa040 [AT] gmail.com

⁴Department of Pharmacology, Faculty of Pharmaceutical Sciences
University of Jos, Plateau State Nigeria
Email: samuelede364 [AT] gmail.com

⁵Department of Human Anatomy, Faculty of Basic Medical Sciences
Federal University Lafia, Nasarawa State Nigeria
Email: alishugaba [AT] yahoo.com

⁶Department of Human Anatomy, Faculty of Basic Medical Sciences
University of Jos, Plateau State Nigeria
Email: eoekwere [AT] gmail.com

ABSTRACT – *This study investigates the protective effects of Nigella sativa oil (NSO) against cyclophosphamide-induced ovarian toxicity in female rats. The research evaluates histological changes in ovarian tissue and their correlation with serum gonadotropin levels. Female rats were assigned to three experimental groups treated with cyclophosphamide, NS-Oil, or a combination, alongside a control group. Ovarian histology was assessed for structural integrity, follicular development, and toxicity, while serum estradiol and progesterone levels were measured. Cyclophosphamide administration significantly disrupted ovarian architecture, reducing follicle count and inducing follicular degeneration. NS-Oil treatment demonstrated protective effects, restoring normal ovarian histology and improving gonadotropin levels. Although body weight showed no significant difference among groups, ovarian weight decreased in cyclophosphamide-treated rats ($p < 0.05$) but increased dose-dependently with NS-Oil treatment ($p < 0.05$). Cyclophosphamide significantly reduced ovarian cortex and medulla volumes ($p < 0.05$), while NS-Oil treatment reversed these effects ($p < 0.05$). Pre-antral and antral follicle counts declined with cyclophosphamide but increased following NS-Oil administration ($p < 0.05$). Atretic follicle counts were lower in NS-Oil-treated groups ($p < 0.05$). NS-Oil also enhanced follicular diameters affected by cyclophosphamide ($p < 0.05$). These findings suggest that NS-Oil may mitigate ovarian toxicity caused by cyclophosphamide, offering a potential therapeutic approach to preserve reproductive health during chemotherapy. Further research is needed to elucidate its mechanisms and clinical applicability.*

Keywords--- *Nigella sativa*, Reproduction, Histomorphology, Ovarian toxicity

1. INTRODUCTION

New study conducted by the WHO, 2023 indicated that there are significant number of people affected by infertility globally, making around 17.5% of the adult population. This is roughly estimated to be 1 in 6 couples worldwide experience infertility, highlighting the burden of infertility, and the need to sought for more studies to provide alternative approaches that is accessible and affordable fertility care for those in need¹.

Infertility, defined as the failure to conceive a recognized pregnancy after 12 months of unprotected intercourse, carries significant personal, societal and financial consequences². Infertility remains a highly prevalent condition worldwide, occurring in 8-12% of couples³. However, the incidence varies from one region of the world to the other, being highest in the so-called infertility belt of Africa that including south Asia, sub-Saharan Africa, the Middle East and North Africa, Central and Eastern Europe and Central Asia³. In contrast to an average prevalence rate of 10-15% in the developed countries⁴, the prevalence of infertility has been notably highly variable in sub-Saharan Africa ranging from 20-46%. This has been attributed to high rate of sexually transmitted diseases, complications of unsafe abortions, and puerperal pelvic infections⁴. About 30% of infertility is due to female problems, 30% to male problems, and 30% to combined male/female problems, while in 10%, there is no recognizable cause⁵.

Nigella sativa, commonly known as black cumin, is an annual flowering plant belonging to the Ranunculaceae family. It is native to the Mediterranean Basin but has naturalized in various regions, including Southern Europe, Southeast Asia, Northern Africa, Eastern Africa, and North America⁶. The plant is characterized by a slender stem (20-30 cm), linear leaves, delicate white or purplish flowers with 5-10 petals, and large capsules containing numerous small, sharp black seeds⁷. These seeds have been extensively studied for their phytochemical properties, which include essential oils⁸, alkaloids⁹, fatty acids¹⁰, sterols¹¹, saponins¹², tannins¹³, flavonoids¹⁴, natural organic acids¹⁵, vitamins and minerals¹⁶.

Nigella sativa has a long history of traditional medicinal use in Asia for treating various ailments, such as headaches, infertility, fever, migraines, diabetes, hypertension, inflammation, and cancer^{17, 18}. The seeds are known for their antimicrobial, analgesic, diuretic, hepatoprotective, cardioprotective, and neuroprotective properties. From the pharmacological stand point, the whole plant is employed in Asia to treat several diseases including headache, infertility, fever, migraine, diabetes, hypertension, inflammations and cancer¹⁹. Indeed, the seeds are used in Asia to exert many properties including the antimicrobial, analgesic, diuretic, antioxytotoxic, antinociceptive, hepatoprotective, cardioprotective and neuroprotective ones²⁰.

Fertility study of *Nigella sativa* seeds has shown to increase the weight of reproductive organs, sperm motility and count in cauda epididimides and testicular ducts in the male rats, while there was an increase in the number of female pregnant rats²¹. *Nigella sativa* oil increased the secretion of sexual hormones, which improved hepatic enzyme protein synthesis, white blood cell count, and blood cholesterol concentration²².

Problem Statement/Justification

Despite advancements in healthcare services, medical interventions, and assisted reproductive technologies in gametes and embryo manipulations, such as Invitro fertilization (IVF), Infertility still remains a major problem that affects most couples all over the world. In Africa, impaired fertility is a well-known public health issue²³. It is a common source of marital problem and disharmony, and is more aggravated in the African society. Several studies indicate that infertility is the most frequent reason for gynecological consultation in Nigeria²⁴. More than 50% of gynecological caseloads are as a result of infertility²⁵, and over 80% of laparoscopic investigations are as a result of infertility²⁸. The International Family planning perspectives in 2017 declared that about 33% of women aged 20-44years suffer from one form of infertility in Nigeria. A substantial proportion of all those affected in developing countries, live in extreme poverty and cannot afford established medical institutions, equipped to address cases of infertility. They resort to the use of traditional folklore medicine, which employs the use of plants and its phytochemical compounds, and is readily available, accessible and very much affordable. This study was aimed at investigating the cyclical changes and histomorphological effects of *Nigella sativa* oil on the ovaries, reproductive tract, hypothalamo-pituitary-ovarian axis, plasma gonadotropins and folliculogenesis in adult female wistar rats.

2. METHODS

2.1 Experimental Design:

The research protocols were approved by the Institutional Animal Care and Use (IACU) protocol, University of Jos, issued by the ethical committee animal experimental unit with approval number: UJ/FPS/F17-00379.

2.2 Experimental Animals:

Twenty-four adults female Wistar rats weighing 120-140g were obtained from the Livestock section of the School of Veterinary and Medical Laboratory Sciences at the National Veterinary Research Institute (NVRI) in Vom. All experiments were conducted in the Vivarium of the same institution. The rats were housed in plastic cages under standard laboratory conditions, including room temperature, humidity, and a 12-hour light/dark cycle. The animals' cages were cleaned daily, and they were fed a standard laboratory diet (Grand Cereal Animal Feeds, Vom) and provided drinking water ad libitum. The rats were acclimated to this environment for two weeks before the study began. All animal care and procedures complied with the guidelines of the institute's ethical committee for medical research. The rats were then randomly divided into four groups of six animals each. *Nigella sativa* oil was administered orally using a gastric tube.

2.3 Plant Acquisition and Preparation:

Nigella sativa seeds were purchased from a commercial shop (Makkah and Madina shop) in Jos metropolis Plateau State, Nigeria. The plant seeds were identified by Mr. Bulus, a taxonomist, Department of Plant Sciences University of Jos, and authenticated by Mr. J.J. Azilla a Botanist from the Federal School of Forestry, Jos. It was given an herbarium voucher number of 0768. The seeds were prepared for phytochemical and elemental analysis, and also oil extraction using Soxhlet method was carried out at the Pharmacognosy Laboratory, Department of Pharmacognosy and Traditional Medicine, Faculty of Pharmaceutical Sciences, University of Jos, Nigeria. Cyclophosphamide (Cycloxan 500mg), manufactured by Zydus Cellexa (Cedila Healthcare Ltd No. 2 ALEAP Industrial Estate, Medical District-500 090 Telengana State, India) was obtained from Le Med Pharmacy, Bauchi Road Jos, Plateau State, Nigeria.

2.4 Induction of Gonadal-toxicity:

Cyclophosphamide is an alkylating agent which has been used to treat the different malignant and nonmalignant diseases, it may have some adverse effects including gastro-intestinal disorder, mutagenesis, pulmonary fibrosis, kidney infection, impaired fertility and could induce premature ovarian failure^{30, 31}. In this study, Cyclophosphamide is selected to induce ovarian failure as animal infertile models. Therefore, the aim of this study is evaluation of the effect of *Nigella sativa* oil on ovarian structures in Cy-induced ovarian failure in female rats.

Histomorphological Studies of Ovaries and Reproductive Tract:

Tissues were fixed in Bouin's solution and dehydrated using graded alcohol concentrations. This method involved dehydration of tissue in 70% alcohol, 90% alcohol, 95% alcohol, and absolute alcohol, each stage lasting for 30minutes. The use of ascending concentration of alcohol is to prevent the rapid dehydration of tissue thereby causing structural damage to the tissue. The dehydrated tissue is cleared in two changes of chloroform for 120minutes each. The clearing is to remove the opacity caused by the dehydrating agent, and make the tissue transparent. The tissue is then infiltrated by immersion into molten paraffin for a period of 30minutes. The tissue was then embedded into molten paraffin wax and allowed to solidify. The embedded tissue was blocked in a rectangular block and sectioned using the rotary microtome at 5µm per section. The tissue sections were allowed to float in water bath at 30°C to help the spreading of the tissue ribbons. Clean slides were used to pick the tissues from the warm water bath. The slides were left to dry and later stained using H&E, and PAS stains

Limitations of the Study

While the present study demonstrated the protective effects of *Nigella sativa* oil (NSO) against cyclophosphamide-induced ovarian toxicity through histological and hormonal assessments, the precise molecular pathways underlying these effects were not explored. Additionally, inflammatory cytokines, including tumor necrosis factor-alpha (TNF-α) and interleukin-6 (IL-6),

which are known mediators of chemotherapy-induced ovarian damage, were not assessed. Furthermore, apoptotic markers such as caspase-3 and Bcl-2, which could provide evidence of NS-Oil's role in follicular cell survival, were not investigated. Future studies incorporating molecular analyses are necessary to elucidate the mechanistic pathways through which NSO exerts its protective effects on ovarian function.

Table 1. Experimental Group Design

Experimental groups	Treatment/rats (induced infertility)
Group 1	Positive Control
Group 2	Negative Control
Group 3	200mg/kg NS oil + 0.5 mg/kg Cyclophosphamide
Group 4	400mg/kg NS oil + 0.5 mg/kg Cyclophosphamide
Group 5	800mg/kg NS oil + 0.5 mg/kg Cyclophosphamide

3. RESULTS

Table 2. Percentage of rats at different estrus phases from different dose levels of *Nigella sativa* oil. Mean \pm S.E.M

Doses (mg/kg body weight)	Percentage (Mean \pm S.E.M) of rats at different phases of the Estrous cycle			
	Proestrus	Estrous	Metestrus	Diestrus
Group I (Positive control)	27.06 \pm 4.3	25.67 \pm 4.2	18.53 \pm 2.3	31.23 \pm 2.8
Group II (Negative control)	37.03 \pm 4.7	27.30 \pm 4.7	24.23 \pm 2.7	17.36 \pm 3.1
Group III (200mg/kg)	47.35 \pm 5.3	31.34 \pm 5.3	18.53 \pm 7.3	7.52 \pm 3.2
Group IV (400mg/kg)	52.07 \pm 2.3	36.07 \pm 3.2	15.63 \pm 2.3	7.20 \pm 5.6
Group V (800mg/kg)	50.07 \pm 3.2	37.42 \pm 4.3	17.73 \pm 3.6	9.03 \pm 1.8

Mean with different superscript in a row are significantly different when compared with the control group at $p < 0.05$

Table 3. Weight of the Uterus, fallopian tubes, and Ovaries of Cyclophosphamide induced gonadotoxic rats administrated with *Nigella sativa* oil for 21 days across the groups

Groups	Estradiol Mean(pg/ml) \pm SEM	Progesterone Mean(pg/ml) \pm SEM
Group I (positive control)	7.91 \pm 0.13 ^{ab}	6.31 \pm 0.13 ^b
Group II (negative control)	6.03 \pm 0.17	5.87 \pm 0.14
Group III (200 mg/kg)	7.41 \pm 0.14	5.61 \pm 0.09
Group IV (400 mg/kg)	7.70 \pm 0.13	6.20 \pm 0.10
Group V (800 mg/kg)	6.76 \pm 0.14 ^a	7.46 \pm 0.14 ^{ab}

SEM = Standard Error in Mean

^{ab} = Statistically significant

Mean with different superscript in a row are significantly different when compared with the control group at $p < 0.05$

Table4: Mean serum level of Estradiol and Progesterone cyclophosphamide induced gonadotoxic rats administered with *Nigella sativa* oil for 21 days

Group	Weight of Uterus, fallopian tubes, and Ovaries Mean weight (g) ± SEM	Body weight/Organ ratio Mean weight (g) ± SEM	P- value
Group I (Positive control)	0.58 ± 0.04	231 ± 5.3	0.043
Group II (Negative control)	0.54 ± 0.04	219 ± 4.7 ^{ab}	0.042
Group III (200mg/kg)	0.56 ± 0.04	227 ± 5.1	0.047
Group IV (400mg/kg)	0.56 ± 0.04	232 ± 4.9	0.046
Group V (800mg/kg)	0.57 ± 0.06	233 ± 4.0 ^{ab}	0.057

P<0.05; S.E.M = standard error in mean; ab = level of significant

Table 5. Effects of *Nigella sativa* oil on the total volume of ovary, cortex and medulla of Cyclophosphamide induced gonadotoxicity rats.

Groups	Cortex volume (µm3)	Medulla volume (µm3)	Total ovary volume (µm3)
Group I (Positive control)	12.9 ± 2.33	8.01 ± 1.23	20.93 ± 2.1
Group II (Negative control)	10.4 ± 2.11 ^a	6.44 ± 0.50 ^a	16.79 ± 2.32 ^a
Group III (200mg/kg)	12.48 ± 1.46 ^b	6.74 ± 1.11 ^b	19.22 ± 1.86 ^b
Group IV (400mg/kg)	12.77 ± 1.99	8.02 ± 0.89	20.80 ± 1.72 ^a
Group V (800mg/kg)	12.53 ± 1.37	8.41 ± 1.72	19.72 ± 1.62

The data are expressed as the Mean ± Standard Error in Mean (S.E.M) p <0.05

Table 6. Mean volume of oocyte (µm3) in different types of follicles in different groups of rats 21 days after treatment with *Nigella sativa* oil on induced Cyclophosphamide gonadotoxic rats. Groups (n=6)

Oocyte volume (µm3)				
Group	Primordial	Primary	Secondary	Antral
Group I (Positive Control)	1235.93 ± 92.1 ^a	2582.08 ± 241.9 ^a	4257.44 ± 346.2 ^a	1248.01 ± 254.7 ^a
Group II (Negative Control)	1252.24 ± 82.1 ^a	3579.61 ± 272.7 ^a	4196.33 ± 318.7 ^a	1752.92 ± 481.3 ^b
Group III (200mg/kg)	1357.70 ± 70.7 ^a	3595.00 ± 253.5 ^a	5775.24 ± 404.0 ^{ab}	1827.00 ± 478.1 ^a
Group IV (400mg/kg)	1435.43 ± 71.2 ^a	3730.09 ± 335.5 ^{ab}	4587.86 ± 438.5 ^a	1594.73 ± 560.1 ^a
Group V (800mg/kg)	1323.43 ± 50.3 ^a	3570.09 ± 327.5 ^a	4857.86 ± 428.5 ^{ab}	1587.86 ± 310.5 ^a

Means with different code letters in each column differ significantly from each other (p < 0.001).

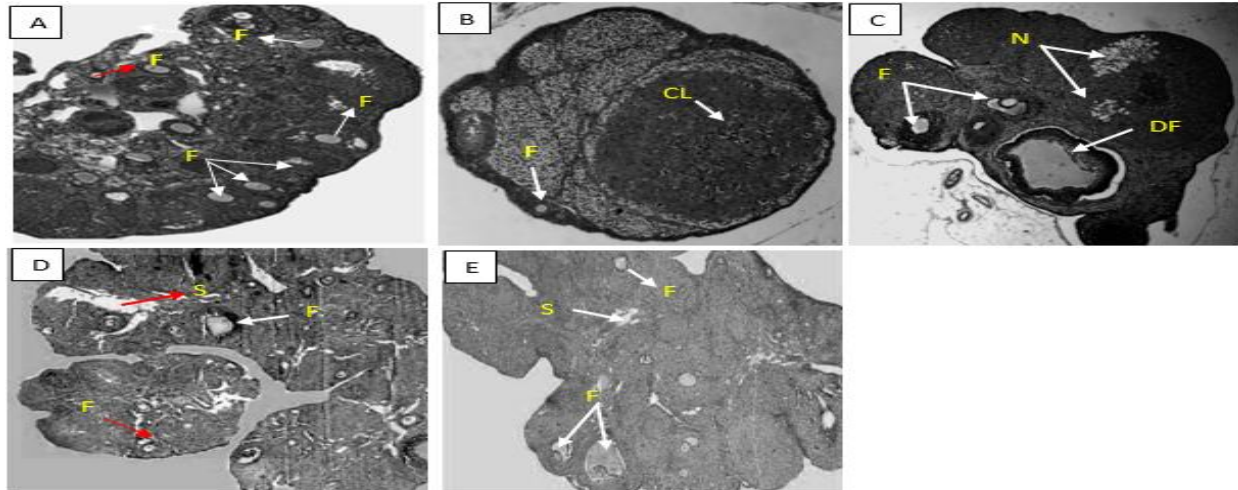


Figure 1: Photomicrograph of Cyclophosphamide induced Ovaries in all experimental groups, after treatment with different concentrations of NS-oil for 21 days. Magnification x 40

- A. Group 1 (Positive Control): Ovaries at different developmental stages, with normal histological architecture
- B. Group 2. (Negative Control): Shows diminished number of developing follicles and increased number of atretic follicles
- C. Group 3. (200mg/kg NS-oil + 0.5 mg/kg Cy): Shows distribution of developing follicles, degenerating follicle and a balloon tissue necrosis on the ovarian surface
- D. Group 4 (400mg/kg NS-oil + 0.5 mg/kg Cy): Presence of developing primary follicles and distorted ovarian stroma
- E. Group 5 (800mg/kg NS-oil + 0.5 mg/kg Cy): Shows ovary with follicles at different stage of development, and mild necrosis on the surface

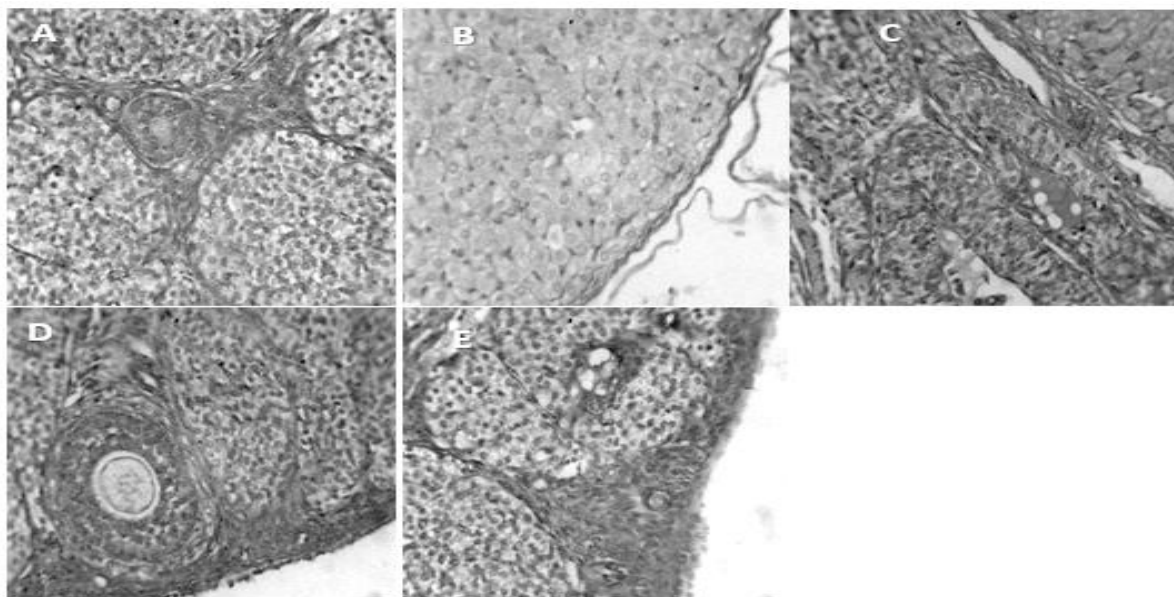


Figure 2. Micrographs showing cross sections of Ovarian Stroma in all treatment groups and the controls. Magnification x100

- A. Group 1. (Positive Control)
- B. Group 2. (Negative Control)
- C. Group 3. (200mg/kg NS-oil + 0.5mg/kg Cy)
- D. Group 4. (400mg/kg NS-oil + 0.5mg/kg Cy)
- E. Group 5. (800mg/kg NS-oil + 0.5mg/kg Cy)

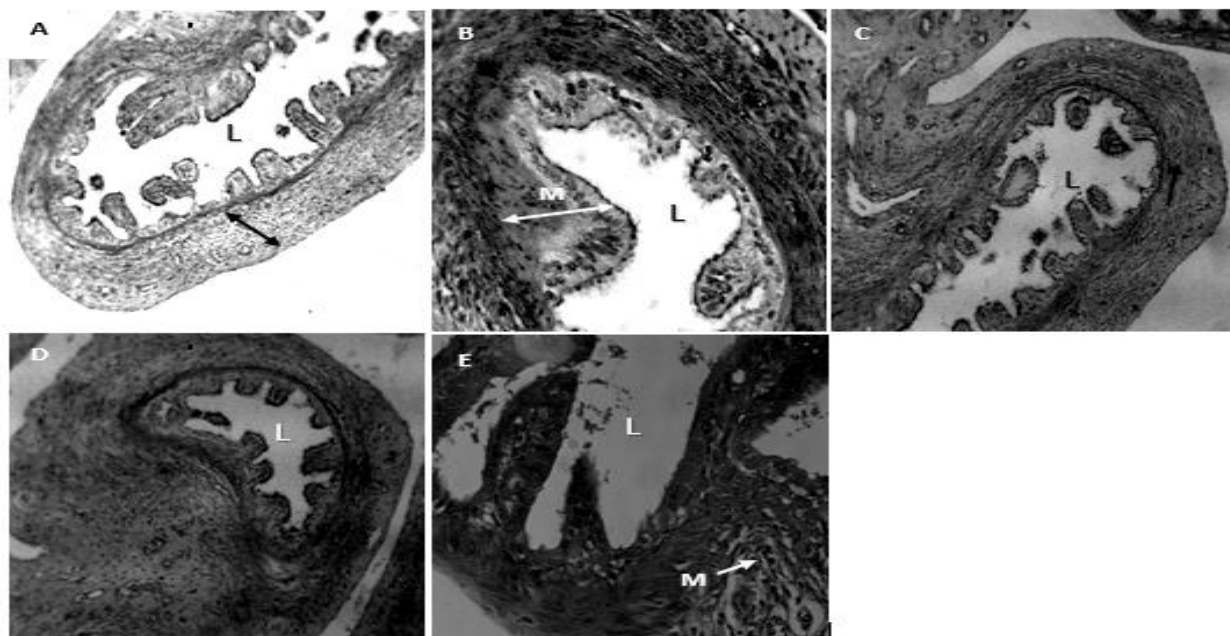


Figure 3: Photomicrographs showing Uterine tubes of experimental rats exposed to 0.5mg/kg Cyclophosphamide and treated with NS-oil of different concentrations for 21days. Magnification x 40

- A. Group 1 (Positive Control): Normal appearance of uterine mucous membrane with branching folds, and central lumen (L).
- B. Group 2. (Negative Control): Shows increased activity of the mucous membrane of the uterine tube
- C. Group 3. (200mg/kg NS-oil + 0.5mg/kg Cy), also shows decrease mucous activity, the muscularis mucosa also shows decrease in diameter
- D. Group 4. (400mg/kg NS-oil + 0.5mg/kg Cy), Narrowing of the luminal diameter can be seen in this group
- E. Group 5. (800mg/kg NS-oil + 0.5mg/kg Cy). The uterine tube is less intact and numerous distortions can be seen. There is also and increased proliferation of the mucous membrane

Body and ovarian weight (Table 3). shows that the mean body weight of the animals in all groups had no significant difference. Ovarian weight reduced in the Cyclophosphamide induced experimental groups compared with the control group ($p < 0.05$), but ovarian weight increased after treatment in the *Nigella sativa* group compared with the control in dose dependent manner ($p < 0.05$). There was no significant difference in these parameters between the experimental and control groups (Table 3).

Volume of the cortex, medulla and total volume of the ovary, and also cortex and medulla volume were reduced in the Cy experimental groups compared with the control group ($p < 0.05$). However, the total volume of the ovary and its cortex were increased after treatment with NS-oil compared with the control experimental groups ($p < 0.05$). There was no significant difference in these parameters between the NS-oil experimental and control groups (Table 5). Number of pre-antral, antral, and atretic follicles Number of pre-antral and antral follicles were reduced in the Cy experimental groups compared with the vehicle group ($p < 0.05$), but pre-antral and antral follicles increased after treatment with NS-oil compared with the Cy experimental groups ($p < 0.05$).

The number of atretic follicles decreased in Cy and NS-oil treated rats compared with the vehicle group ($p < 0.05$). There was no significant difference in these parameters between the NS-oil experimental and vehicle groups (Table 6). Diameter of pre-antral and antral follicles reduced in the Cy experimental groups compared with the vehicle group ($p < 0.001$). The diameter of pre-antral and antral follicle increased after treatment with NS-oil compared with the Cy experimental groups ($p < 0.05$). There was no significant difference in these parameters between the NS-oil experimental and vehicle groups (Table 6).

Diameter of follicular cells in pre-antral and antral follicles Diameter of follicular cells in pre-antral and antral follicle were reduced in the Cy experimental groups compared with the vehicle group ($p < 0.01$) and diameter of follicular cells in pre-antral

and antral follicle were increased after treatment with NS-oil related to the Cy experimental groups ($p < 0.05$) and decreased compared with the vehicle.

Diameter of oocyte in pre-antral and antral follicles Diameter of oocyte in pre-antral follicle was reduced in Cy and NS-oil experimental groups compared with the vehicle group ($p < 0.05$). Diameter of oocyte in antral follicle was reduced in the Cy and NS-oil experimental groups compared with the vehicle group ($p < 0.05$). There was no significant difference in this parameter between the other experimental groups (Table 6)

4. DISCUSSION

The rat has a brief reproductive cycle of 4 to 5 days in phases, which makes it excellent for reproductive investigations^{37,38}. The release of gonadotropin-releasing hormone from the brain, gonadotropins from the pituitary, and sex hormones from the gonads all impact the estrous cycle in sexually developed female animals^{39,40}. Plant extracts inhibited the estrous cycle in rats during the diestrus phase, preventing the production of both follicle-stimulating hormone (FSH) and luteinizing hormone (LH)^{41,42}.

The loss of regular cyclicity implies that the ovarian progesterin and estrogen equilibrium has been disrupted. Studies reported that many extracts resulted to lengthened the diestrus stage of the cycle^{43,44}, and sometimes complete abolition of proestrus, with prolonged estrus, and shortened diestrus phases of the cycle following the administration of *Nigella sativa* oil^{45,46}. The results of the current study (Table 2) confirm these previous claims of the ability of some plant extracts to lengthen the diestrus phase of the cycle. Distinct plants have different ways for altering the reproductive cycle, as evidenced by the submissions.

The result in this studies shows that there was an increase in estradiol hormone and progesterin in the rats that were treated with 800mg/kg *Nigella sativa* oil (Table 4), this is in line with several studies of *Nigella sativa* oil on reproduction, showed that it stimulated the secretion of sexual hormones that led to improve synthesis of hepatic enzymes^{47,48}, it increases the thickness of germinal layer of seminiferous tubules significantly^{49,50}, also reverse toxicity effect in rats treated with Lead acetate (100mg/kg), and causes significant enhancement in reproductive function, increases ovarian weight to body ratio, gonadotrophins, and diameter of graafian follicles^{51,52}.

Ovarian failure is distinguished by ovarian atrophy, follicles reduction, and sex hormonal diminution⁵³. Cyclophosphamide is an alkylating agent which induce ovarian failure in animal models^{54,55}. This quantitative study showed that Cy decreased weight and volume of ovary, reduced different follicles number and sex hormones levels and increased atretic follicles compared to vehicle group. Reduction of the medulla volume may be attributed to the rate of blood circulation. suggesting that, reduction of the rate of medulla blood circulation caused to medulla volume diminution. Reduction of cortex volume can be related to follicular number decrement. This result indicates that Cy had the most important effect on the number of pre-antral follicles relative to antral follicle and increase atretic follicle⁵⁶, showed that Cy causes degeneration of ovarian follicle number in mice. Other studies have demonstrated that Cy significantly reduces primordial and primary follicles and increases the numbers of atretic follicles^{57,58}. This result also shows that Cyclophosphamide adversely affect follicular development at all stages of development. This is in line with Epstein⁶⁰, who believed that Cy has negative effect on both resting and dividing cells. Lopez and Luderer (2014) show that Cy leads to misdirection of the diakinesis stage in first meiosis of oögonia^{61,62}. Other researches have demonstrated that Cy inhibits follicular cell division through DNA strand break and inhibit mitotic divisions^{63,64}. Based on this information, the other parameters should be measured to confirm this data. In this research, cyclophosphamide reduced the diameters of different follicles, oocyte and follicular cells relative to vehicle group. Lopez and Luderer (2004) demonstrated that Cy-induced follicular cell apoptosis in rats⁶⁵. Other studies showed that ovarian failure had the most side effect on follicular cells compared with resting follicle^{67,68}. Researches show that, any factor which induces follicle syncope in the primary follicle stage, inhibit follicular cell mitosis, consequently, follicular cell remain only one layer and formation of any new layers of follicular cells are uncommon^{69,70}. It seems reduction of follicular cell in pre-antral follicle prevent germ cell development and decrease the number of pre-antral and antral follicles. In addition, reduction of follicular cells has direct influence on the size of oocyte in growing follicle.

This study also demonstrated that Cy decreased the level of estradiol and progesterone. Follicular cells are the main source of estradiol and progesterone. Destruction of antral and pre-antral follicles have a direct effect on estradiol level⁷¹. Therefore, it seems that change in the number of the growing follicles influences the level of hormones. Studies show that growth factors have an important role in improving the structure and function of ovaries. Different growth factors such as CTGF, EGF, and TGF- β have protective effects on ovarian damage^{72,73,74,75,76}. In this study, it was found that NS oil had a positive effect on the

volume of the ovarian cortex, the number of preantral follicles, and the diameter of the antral follicles. These results also show that NS oil increases the volume of the ovarian cortex, the number of preantral follicles, and the diameter of the antral follicle and its ovule.

Based on these results, it can be proposed that NS-oil accelerates antral folliculogenesis by stimulating follicular cell proliferation and thereby increasing antral follicle diameter. An increase in oocyte diameter in the antral follicles under the influence of growth factors is a common phenomenon, while a decrease in oocyte diameter in the preantral follicles under this condition is an unusual phenomenon. The mechanism of cell size is not fully understood but is determined by external and internal factors^{77, 78}. Zetterberg believe that macromolecular factors such as EGF have a stimulatory effect on DNA synthesis but do not affect cell size⁷⁹. In addition, GDF9 stimulates meiotic division in metaphase I and II and regulates follicle and ovarian size^{80, 81}. In this study, the extract oil of NS-oil had no effect on estradiol and progesterone levels. The follicular cells do not appear to have reached their full development; Therefore, the levels of these hormones have not changed significantly. In this study, we also found that NS-oil administration had no effect on various ovarian structures and functions in normal rats. It appears that the beneficial effects of NS-oil apply only to damaged ovaries and do not affect normal structure. Conclusively, Cyclophosphamide had a more damaging effect on follicular cells than on oocytes in different ovarian follicles. NS-oil had a positive effect on follicle growth and improved ovarian tissue repair.

5. CONCLUSION

The results of this investigation provide light on the intricate interactions that occur between plant extracts of NS oil, in particular and reproductive physiology in rat models, especially when cyclophosphamide (Cy) is present. The study demonstrated that exogenous chemicals and innate physiological processes have a substantial impact on the short estrous cycle in female rats.

Cy treatment caused significant ovarian dysfunction, which included ovarian shrinkage, a drop in the number of follicles, and low sex hormone levels, especially progesterone and estradiol. This is consistent with other research showing that Cy causes ovarian failure by means of follicular cell death and DNA damage, which ultimately results in abnormal folliculogenesis and hormonal abnormalities. These results are supported by the study's quantitative analysis, which shows a notable rise in atretic follicles and a decrease in the volume of the ovarian cortex and medulla, both of which are essential for healthy reproductive function.

On the other hand, NS oil administration showed ovarian structures to be protected, increasing the number of preantral follicles and the diameter of antral follicles, indicating that it may have a role in folliculogenesis and ovarian tissue healing. The fact that NS oil treatment raised estradiol and progesterone levels added to the evidence supporting its effectiveness in promoting ovarian function, especially in impaired situations brought on by Cy.

These findings highlight the significance of comprehending the processes by which NS oil extracts can alter hormone levels and reproductive cycles, laying the groundwork for future therapeutic uses in the treatment of reproductive dysfunctions. Future studies should concentrate on clarifying the precise mechanisms by which *Nigella sativa* oil works to cure ovarian-related diseases and investigating the oil's potential use in clinical settings. All things considered, this research offers insightful information about the dynamics of reproductive health and the possibility that natural substances can lessen the negative effects of synthetic medications like cyclophosphamide.

6. ACKNOWLEDGEMENT

The Authors would like to acknowledge the following:

1. Sa'adu Zungur University Bauchi, Bauchi State Nigeria
2. Tertiary Education Trust (TET) Fund Nigeria
3. Association of Academic Staff Union of Universities (ASUU) Nigeria

7. REFERENCES

1. Abbaspour, N., Hurrell, R., & Kelishadi, R. (2014). Review on iron and its importance for human health. *Journal of Research in Medical Sciences : The Official Journal of Isfahan University of Medical Sciences*, 19(2), 164–174. <https://pubmed.ncbi.nlm.nih.gov/24778671>

2. Abd El Aziz, A. E., el Sayed, N. S., & Mahran, L. G. (2011). Anti-asthmatic and anti-allergic effects of thymoquinone on airway-induced hypersensitivity in experimental animals. *Journal of Applied Pharmaceutical Science*, 1(8), 109–117.
3. Abd Rani, N. Z., Husain, K., & Kumolosasi, E. (2018). Moringa Genus: A Review of Phytochemistry and Pharmacology . In *Frontiers in Pharmacology*. (Vol. <https://www.frontiersin.org/article/10.3389/fphar.2018.00108>
4. Abdelsalam, S. A. E. and E. B. (2018). Some Biological and Pharmacological Effects of the Black Cumin (*Nigella sativa*): A Concise Review. *American Journal of Research Communication*, 6(3). www.usa-journals.com,
5. Abdulrahman, F. T. J. H. M. A. (20 C.E.). The effects of *Nigella sativa* oil administration on some physiological and histological values of reproductive aspects of rats: *The Iraqi Journal of Veterinary Medicine*, 35(2)(50–60). <https://doi.org/10.30539/iraqijvm.v35i2.576>
6. Abeysinghe, D. T., Kumara, K. A. H., Kaushalya, K. A. D., Chandrika, U. G., & Alwis, D. D. D. H. (2021). Phytochemical screening, total polyphenol, flavonoid content, in vitro antioxidant and antibacterial activities of Sri Lankan varieties of *Murraya koenigii* and *Micromelum minutum* leaves. *Heliyon*, 7(7), e07449–e07449. <https://doi.org/10.1016/j.heliyon.2021.e07449>
7. Adaku, A. (2018). *Essential Oil from Nigella sativa Seed Differentially Ameliorates Steroid Genesis , Cellular ATP and Prostate Functions in Anti-Psychotic Drug- Induced Testicular Damage of Rats*. 8(1), 1–9. <https://doi.org/10.4172/2161-0495.1000371>
8. Adewoyin, M., Ibrahim, M., Roszaman, R., Isa, M., Alewi, N., Rafa, A., & Anuar, M. (2017). Male Infertility: The Effect of Natural Antioxidants and Phytocompounds on Seminal Oxidative Stress. *Diseases*, 5(1), 9. <https://doi.org/10.3390/diseases5010009>
9. Aglave, H. (2018). Physiochemical characteristics of sesame seeds. *Journal of Medicinal Plants Studies*, 6(1), 64–66. <http://dx.doi.org/10.1016/j.foodchem.2006.09.008>
10. Ahmad, A., Husain, A., Mujeeb, M., Siddiqui, N. A., & Damanhour, Z. A. (2012). *Physicochemical and phytochemical standardization with HPTLC fingerprinting of Nigella sativa L . seeds*. 1175–1182.
11. Ahmad, A., Mishra, R. K., Vyawahare, A., Kumar, A., Rehman, M. U., Qamar, W., Khan, A. Q., & Khan, R. (2019). Thymoquinone (2-Isopropyl-5-methyl-1, 4-benzoquinone) as a chemopreventive/anticancer agent: Chemistry and biological effects. *Saudi Pharmaceutical Journal*, 27(8), 1113–1126. <https://doi.org/10.1016/j.jsps.2019.09.008>
12. Ahmad, F., Ali, F., Amir, S., Saad, H. H., Wahab, S., Idreesh, M., Ali, M., & Mohan, S. (2020). *Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID- 19 . The COVID-19 resource centre is hosted on Elsevier Connect , the company ' s public news and information . January*.
13. Ahmad, W., Zeenat, F., & Shaiqua, A. (2017). *Therapeutics , Phytochemistry and Pharmacology of an Important Unani Drug Kalonji (Nigella sativa Linn) : A Review THERAPEUTICS , PHYTOCHEMISTRY AND PHARMACOLOGY OF AN IMPORTANT*. July.
14. Ajayi, A. F., & Akhigbe, R. E. (2020). Staging of the estrous cycle and induction of estrus in experimental rodents: an update. *Fertility Research and Practice*, 6(1), 1–15. <https://doi.org/10.1186/s40738-020-00074-3>
15. al Disi, S. S., Anwar, M. A., & Eid, A. H. (2016). Anti-hypertensive herbs and their mechanisms of action: Part I. *Frontiers in Pharmacology*, 6(JAN), 1–24. <https://doi.org/10.3389/fphar.2015.00323>
16. Alanazi, I. O., Benabdelkamel, H., Alfadda, A. A., AlYahya, S. A., Alghamdi, W. M., Aljohi, H. A., Almalik, A., & Masood, A. (2016). Proteomic Analysis of the Protein Expression Profile in the Mature *Nigella sativa* (Black Seed). *Applied Biochemistry and Biotechnology*, 179(7), 1184–1201. <https://doi.org/10.1007/s12010-016-2058-z>
17. Alberts B, Johnson A, Lewis J, et al. (2002). *Molecular Biology of the Cell*. Garland Science. <https://www.ncbi.nlm.nih.gov/books/NBK26940/%0A>
18. Algandaby, M. M. (2021). Quercetin attenuates cisplatin-induced ovarian toxicity in rats: Emphasis on anti-oxidant, anti-inflammatory and anti-apoptotic activities. *Arabian Journal of Chemistry*, 14(7), 103191. <https://doi.org/10.1016/j.arabjc.2021.103191>
19. Ali, B. A. (2001). *on Blood Glucose in Albino Rats*. 242–244.
20. Al-Johar, D., Shinwari, N., Arif, J., Al-Sanea, N., Jabbar, A. A., El-Sayed, R., Mashhour, A., Billedo, G., El-Doush, I., & Al-Saleh, I. (2008). Role of *Nigella sativa* and a number of its antioxidant constituents towards azoxymethane-

- induced genotoxic effects and colon cancer in rats. *Phytotherapy Research*, 22(10), 1311–1323. <https://doi.org/https://doi.org/10.1002/ptr.2487>
21. Al-Mamun, M., & Absar, N. (2019). Major nutritional compositions of black cumin seeds-cultivated in Bangladesh and the physicochemical characteristics of its oil. *International Food Research Journal*, 25, 2634–2639.
22. Alomar, M. Y. (2020). Physiological and histopathological study on the influence of *Ocimum basilicum* leaves extract on thioacetamide-induced nephrotoxicity in male rats. *Saudi Journal of Biological Sciences*, 27(7), 1843–1849. <https://doi.org/https://doi.org/10.1016/j.sjbs.2020.05.034>
23. Amin, B., & Hosseinzadeh, H. (2016). Black Cumin (*Nigella sativa*) and Its Active Constituent, Thymoquinone: An Overview on the Analgesic and Anti-inflammatory Effects. *Planta Medica*, 82(1-2)(8–16). <https://doi.org/10.1055/s-0035-1557838>
24. Amin, B., & Hosseinzadeh, H. (2016). Black Cumin (*Nigella sativa*) and Its Active Constituent, Thymoquinone: An Overview on the Analgesic and Anti-inflammatory Effects. *Planta Medica*, 82(1–2), 8–16. <https://doi.org/10.1055/s-0035-1557838>
25. Andrade, G. M., Collado, M. del, Meirelles, F. V., da Silveira, J. C., & Perecin, F. (2019). Intrafollicular barriers and cellular interactions during ovarian follicle development. *Animal Reproduction*, 16(3), 485–496. <https://doi.org/10.21451/1984-3143-AR2019-0051>
26. Angad, G., Veterinary, D. E. v, & Sciences, A. (2015). *HISTOMORPHOCHEMICAL AND ULTRASTRUCTURAL CHARACTERIZATION OF HYPOTHALAMO-HYPOPHYSEAL-OVARIAN AXIS IN INDIAN BUFFALO (Bubalus bubalis) (Minor Subject : Veterinary Physiology) By Department of Veterinary Anatomy College of Veterinary Science GURU ANGAD D.*
27. Antoniadis, V., Shaheen, S. M., Levizou, E., Shahid, M., Niazi, N. K., Vithanage, M., Ok, Y. S., Bolan, N., & Rinklebe, J. (2019). A critical prospective analysis of the potential toxicity of trace element regulation limits in soils worldwide: Are they protective concerning health risk assessment? - A review. *Environment International*, 127, 819–847. <https://doi.org/https://doi.org/10.1016/j.envint.2019.03.039>
28. Arnal, J., Lenfant, F., Metivier, R., Flouriot, G., Henrion, D., Adlanmerini, M., Fontaine, C., Gourdy, P., Chambon, P., Katzenellenbogen, B., & Katzenellenbogen, J. (2022). *MEMBRANE AND NUCLEAR ESTROGEN RECEPTOR ALPHA ACTIONS: FROM TISSUE SPECIFICITY TO MEDICAL IMPLICATIONS. Figure 2*, 1045–1087. <https://doi.org/10.1152/physrev.00024.2016>
29. *ARN.eBook*. (n.d.).
30. Arroyo, A., Kim, B., & Yeh, J. (2020). Luteinizing Hormone Action in Human Oocyte Maturation and Quality: Signaling Pathways, Regulation, and Clinical Impact. *Reproductive Sciences*, 27(6), 1223–1252. <https://doi.org/10.1007/s43032-019-00137-x>
31. Article, O. (2012). *The enhancing effects of alcoholic extract of Nigella sativa seed on fertility potential, plasma gonadotropins and testosterone in male rats*. 10(4), 355–362.
32. Assi, M. A., Hezmee, M., Noor, M., Farhana Bacheek, N., Ahmad, H., Haron, A. W., Sabri, M., Yusoff, M., & Rajion, M. A. (2016). The Various Effects of *Nigella sativa* on Multiple Body Systems in Human and Animals. *PJSRR Pertanika Journal of Scholarly Research Reviews*, 2(3), 1–19. <http://www.pjsrr.upm.edu.my/>
33. Atanasov, A. G., Zotchev, S. B., Dirsch, V. M., Orhan, I. E., Banach, M., Rollinger, J. M., Barreca, D., Weckwerth, W., Bauer, R., Bayer, E. A., Majeed, M., Bishayee, A., Bochkov, V., Bonn, G. K., Braid, N., Bucar, F., Cifuentes, A., D'Onofrio, G., Bodkin, M., ... Taskforce, the I. N. P. S. (2021). Natural products in drug discovery: advances and opportunities. *Nature Reviews Drug Discovery*, 20(3), 200–216. <https://doi.org/10.1038/s41573-020-00114-z>
34. Atata, J. A., Esievo, K. A. N., Adamu, S., Abdulsalam, H., Avazi, D. O., & Ajadi, A. A. (2019). Haemato-biochemical studies of dogs with haemorrhage-induced dehydration. *Comparative Clinical Pathology*, 28(1), 129–135. <https://doi.org/10.1007/s00580-018-2805-3>
35. Atiku, I. A. (2018). *Cephalic index in relation to academic performance among students of basic medical sciences, bayero university kano, nigeria*. 4(1), 404–415.
36. Badar, A., Kaatabi, H., Bamosa, A., Al-Elq, A., Abou-Hozafa, B., Lebda, F., Alkhadra, A., & Al-Almaie, S. (2017). Effect of *Nigella sativa* supplementation over a one-year period on lipid levels, blood pressure and heart rate in type-

- 2 diabetic patients receiving oral hypoglycemic agents: Nonrandomized clinical trial. *Annals of Saudi Medicine*, 37(1), 56–63. <https://doi.org/10.5144/0256-4947.2017.56>
37. Bagnjuk, K., & Mayerhofer, A. (2019). Human luteinized granulosa cells—a cellular model for the human corpus luteum. *Frontiers in Endocrinology*, 10(JULY), 1–7. <https://doi.org/10.3389/fendo.2019.00452>
38. Bailey, E. J., Chang, A. B., & Thomson, D. (2008). In children with prolonged cough, does treatment with antibiotics have a better effect on cough resolution than no treatment? Part B: Clinical commentary. *Paediatrics and Child Health*, 13(6), 514. <https://doi.org/10.1093/pch/13.6.514>
39. Bakathir, H. A., & Abbas, N. A. (2011). Detection of the antibacterial effect of *Nigella sativa* ground seeds with water. *African Journal of Traditional, Complementary and Alternative Medicines*, 8(2), 159–164. <https://doi.org/10.4314/ajtcam.v8i2.63203>
40. Balali-Mood, M., Naseri, K., Tahergorabi, Z., Khazdair, M. R., & Sadeghi, M. (2021). Toxic Mechanisms of Five Heavy Metals: Mercury, Lead, Chromium, Cadmium, and Arsenic. In *Frontiers in Pharmacology* (Vol. 12). <https://www.frontiersin.org/article/10.3389/fphar.2021.643972>
41. Bashir, M. U., & Qureshi, H. J. (2010). No Title Analgesic effect of *Nigella sativa* seeds extract on experimentally induced pain in albino mice. *Journal of the College of Physicians and Surgeons--Pakistan, JCPSP*, 20((464–467).
42. Benefits, H., Pharmacology, M., Dash, R., Sikder, M. H., Rahman, S., Timalisina, B., & Munni, Y. A. (2021). Black Cumin (*Nigella sativa* L.): A Comprehensive Review on Phytochemistry, Health Benefits, Molecular Pharmacology, and Safety. *Nutrients*.
43. Bieberich, E. (2014). Synthesis, Processing, and Function of N-glycans in N-glycoproteins. *Advances in Neurobiology*, 9, 47–70. https://doi.org/10.1007/978-1-4939-1154-7_3
44. Blaik, P. (2013). No 主観的健康感を中心とした在宅高齢者における健康関連指標に関する共分散構造分析 Title. *Gospodarka Materialowa i Logistyka*, 26(4), 185–197.
45. Blanc, L., & Wolfe, L. C. (2016). Chapter 9 - General Considerations of Hemolytic Diseases, Red Cell Membrane, and Enzyme Defects. In P. Lanzkowsky, J. M. Lipton, & J. D. Fish (Eds.), *Lanzkowsky's Manual of Pediatric Hematology and Oncology (Sixth Edition)* (Sixth Edit, pp. 134–158). Academic Press. <https://doi.org/https://doi.org/10.1016/B978-0-12-801368-7.00009-0>
46. Boskabady, M. H., Keyhanmanesh, R., Khamneh, S., & Ebrahimi, M. A. (2011). The effect of *Nigella sativa* extract on tracheal responsiveness and lung inflammation in valbuminsensitized guinea pigs. *Clinics*, 66(5), 879–887. <https://doi.org/10.1590/S1807-59322011000500027>
47. Bridgewater, P., Upadhyaya, S., Poudyal, B., Kunwar, R. M., Bussmann, R. W., & Paniagua-Zambrana, N. Y. (2020). *Nigella sativa* L. *Ranunculaceae* BT - *Ethnobotany of the Himalayas* (R. Kunwar, H. Sher, & R. W. Bussmann, Eds.; pp. 1–10). Springer International Publishing. https://doi.org/10.1007/978-3-030-45597-2_162-1
48. Brunt, V. E., Miner, J. A., Meendering, J. R., Kaplan, P. F., & Minson, C. T. (2012). *Cutaneous Thermal Hyperemia But Not Reactive Hyperemia*. 18(5), 347–355. <https://doi.org/10.1111/j.1549-8719.2011.00095.x.17->
49. Bruyn, G. W. (1989). Human central nervous system. *Journal of the Neurological Sciences*, 92(1), 117. [https://doi.org/10.1016/0022-510x\(89\)90181-0](https://doi.org/10.1016/0022-510x(89)90181-0)
50. BUREAU OF PUBLIC PROCUREMENT BIDDERS ' CORRESPONDENCE DETAILS TEMPLATE Ministry : Name of Procuring Entity : Title of Procurement : (n.d.).
51. Butt, M. S., & Sultan, M. T. (2010). *Nigella sativa : Reduces the Risk of Various Maladies Nigella sativa : Reduces the Risk*. 8398. <https://doi.org/10.1080/10408390902768797>
52. Caligioni, C. S. (2009). Assessing Reproductive Status/Stages in Mice. *Current Protocols in Neuroscience*, 48(1), A.41.1-A.41.8. <https://doi.org/https://doi.org/10.1002/0471142301.nsa04is48>
53. Campinas, U. E. de, & Campinas, U. E. de. (2002). *DETERMINATION OF THE ESTROUS CYCLE PHASES OF RATS : SOME HELPFUL CONSIDERATIONS*. 62, 609–614.
54. Casarini, L., & Crépieux, P. (2019). Molecular Mechanisms of Action of FSH. *Frontiers in Endocrinology*, 10. <https://doi.org/10.3389/fendo.2019.00305>
55. Castellini, C., Mattioli, S., Signorini, C., Cotozzolo, E., Noto, D., Moretti, E., Brecchia, G., Dal Bosco, A., Belmonte, G., Durand, T., De Felice, C., & Collodel, G. (2019). Effect of Dietary n-3 Source on Rabbit Male Reproduction. *Oxidative Medicine and Cellular Longevity*, 3279670. <https://doi.org/10.1155/2019/3279670>

56. Chaklader, M. R., Fotedar, R., Howieson, J., Siddik, M. A. B., & Foysal, M. J. (2020). The ameliorative effects of various fish protein hydrolysates in poultry by-product meal based diets on muscle quality, serum biochemistry and immunity in juvenile barramundi, *Lates calcarifer*. *Fish & Shellfish Immunology*, 104, 567–578. <https://doi.org/https://doi.org/10.1016/j.fsi.2020.06.014>
57. Chassagne, F., Samarakoon, T., Porras, G., Lyles, J. T., Dettweiler, M., Marquez, L., Salam, A. M., Shabih, S., Farrokhi, D. R., & Quave, C. L. (2021). A Systematic Review of Plants With Antibacterial Activities: A Taxonomic and Phylogenetic Perspective. *Frontiers in Pharmacology*, 11. <https://doi.org/10.3389/fphar.2020.586548>
58. Chauvin, S., Cohen-Tannoudji, J., & Guigon, C. J. (2022). Estradiol Signaling at the Heart of Folliculogenesis: Its Potential Deregulation in Human Ovarian Pathologies. *International Journal of Molecular Sciences*, 23(1). <https://doi.org/10.3390/ijms23010512>
59. ChiauMingj, Md. S. A. A. HingGohefZannatUrbigMd. M. R. (2021). A review of ethnobotany, phytochemistry, antimicrobial pharmacology and toxicology of *Nigella sativa* L. *Biomedicine & Pharmacotherapy*, Volume 143(November 2021, 112182).
60. Choi, Y. J., Kim, N. N., Habibi, H. R., & Choi, C. Y. (2016). Effects of gonadotropin inhibitory hormone or gonadotropin-releasing hormone on reproduction-related genes in the protandrous cinnamon clownfish, *Amphiprion melanopus*. *General and Comparative Endocrinology*, 235, 89–99. <https://doi.org/https://doi.org/10.1016/j.ygcen.2016.06.010>
61. Choudhury, H., Pandey, M., Hua, C. K., Mun, C. S., Jing, J. K., Kong, L., Ern, L. Y., Ashraf, N. A., Kit, S. W., Yee, T. S., Pichika, M. R., Gorain, B., & Kesharwani, P. (2018). An update on natural compounds in the remedy of diabetes mellitus: A systematic review. *Journal of Traditional and Complementary Medicine*, 8(3), 361–376. <https://doi.org/10.1016/j.jtcme.2017.08.012>
62. Chu, Y. L., Xu, Y. R., Yang, W. X., & Sun, Y. (2018). *Aging-V10I3-101391 (1)*. 10(3), 305–321.
63. Clark, A. R., & Stokes, Y. M. (2011). Follicle structure influences the availability of oxygen to the oocyte in antral follicles. *Computational and Mathematical Methods in Medicine*, 2011. <https://doi.org/10.1155/2011/287186>
64. Clarke, H. J. (2018). Regulation of germ cell development by intercellular signaling in the mammalian ovarian follicle. *Wiley Interdisciplinary Reviews: Developmental Biology*, 7(1), 1–33. <https://doi.org/10.1002/wdev.294>
65. Comish, P. B., Drumond, A. L., Kinnell, H. L., Anderson, R. A., Matin, A., Meistrich, M. L., & Shetty, G. (2014). Fetal cyclophosphamide exposure induces testicular cancer and reduced spermatogenesis and ovarian follicle numbers in mice. *PLoS ONE*, 9(4). <https://doi.org/10.1371/journal.pone.0093311>
66. Contreras-Zentella, M. L., & Hernández-Muñoz, R. (2016). Is Liver Enzyme Release Really Associated with Cell Necrosis Induced by Oxidant Stress? *Oxidative Medicine and Cellular Longevity*, 2016, 3529149. <https://doi.org/10.1155/2016/3529149>
67. Coskun, D., Britto, D. T., Shi, W., & Kronzucker, H. J. (2017). How Plant Root Exudates Shape the Nitrogen Cycle. *Trends in Plant Science*, 22(8), 661–673. <https://doi.org/https://doi.org/10.1016/j.tplants.2017.05.004>
68. Cruz, G., Fernandois, D., & Paredes, A. H. (2017). Ovarian function and reproductive senescence in the rat: Role of ovarian sympathetic innervation. *Reproduction*, 153(2), R59–R68. <https://doi.org/10.1530/REP-16-0117>
69. da Broi, M. G., Giorgi, V. S. I., Wang, F., Keefe, D. L., Albertini, D., & Navarro, P. A. (2018a). Influence of follicular fluid and cumulus cells on oocyte quality: clinical implications. *Journal of Assisted Reproduction and Genetics*, 35(5), 735–751. <https://doi.org/10.1007/s10815-018-1143-3>
70. da Broi, M. G., Giorgi, V. S. I., Wang, F., Keefe, D. L., Albertini, D., & Navarro, P. A. (2018b). Influence of follicular fluid and cumulus cells on oocyte quality: Clinical implications. *Journal of Assisted Reproduction and Genetics*, 35(5), 735–751. <https://doi.org/10.1007/s10815-018-1143-3>
71. Darkwah, W. K., Kadri, A., Adormaa, B. B., & Aidoo, G. (2018). Cephalometric study of the relationship between facial morphology and ethnicity: Review article. *Translational Research in Anatomy*, 12(September), 20–24. <https://doi.org/10.1016/j.tria.2018.07.001>
72. David Lazer, Ryan Kennedy, Gary King, & Vespignani Alessandro. (2014). The Parable of Google Flu: Traps in Big Data Analysis. *Science*, 343(March), 1203–1205. www.sciencemag.org/SCIENCEVOL34314MARCH2014
73. *Department Of General Histology General Embriology Introduction*. (n.d.).

74. Dey, S., Samanta, P., Pal, S., Mukherjee, A. K., Kole, D., & Ghosh, A. R. (2016). Integrative assessment of biomarker responses in teleostean fishes exposed to glyphosate-based herbicide (Excel Mera 71). *Emerging Contaminants*, 2(4), 191–203. <https://doi.org/https://doi.org/10.1016/j.emcon.2016.12.002>
75. Dharmalingam, K., Birdi, A., Tomo, S., Sreenivasulu, K., Charan, J., Yadav, D., Purohit, P., & Sharma, P. (2021). Trace Elements as Immunoregulators in SARS-CoV-2 and Other Viral Infections. *Indian Journal of Clinical Biochemistry : IJCB*, 36(4), 416–426. <https://doi.org/10.1007/s12291-021-00961-6>
76. Diederich, L., Iv, T. C. S. K., Kuhn, V., & Kramer, C. M. (2017). *Red Blood Cell Function and Dysfunction* : 26(13), 718–742. <https://doi.org/10.1089/ars.2016.6954>
77. Dollah, M. A., Parhizkar, S., Latiff, L. A., & bin Hassan, M. H. (2013). Toxicity effect of *Nigella sativa* on the liver function of rats. *Advanced Pharmaceutical Bulletin*, 3(1), 97–102. <https://doi.org/10.5681/apb.2013.016>
78. Dubey, P. N., Singh, B., Mishra, B. K., Kant, K., & Solanki, R. K. (2016). *Nigella (Nigella sativa)*: A high value seed spice with immense medicinal potential. *Indian Journal of Agricultural Sciences*, 86(8), 967–979.
79. Ebrahimi, M., & Akbari Asbagh, F. (2011). Pathogenesis and causes of premature ovarian failure: an update. *International Journal of Fertility & Sterility*, 5(2), 54–65.
80. *EFFECT ON REPRODUCTIVE SYSTEM*. (n.d.).
81. Effenberger, K., Breyer, S., & Schobert, R. (2010). *Terpene Conjugates of the Nigella sativa Seed-Oil Constituent Thymoquinone with Enhanced Efficacy in Cancer Cells*. 7, 129–139.
82. Eid, A. M., Elmarzugi, N. A., Ayyash, L. M. A., Sawafta, M. N., & Daana, H. I. (2017). *A Review on the Cosmeceutical and External Applications of Nigella sativa*. 2017.
83. Eleawa, S. M., Alkhateeb, M. A., Alhashem, F. H., Bin-Jaliah, I., Sakr, H. F., Elrefaey, H. M., Elkarib, A. O., Alessa, R. M., Haidara, M. A., Shatoor, A. S., & Khalil, M. A. (2014). Resveratrol reverses cadmium chloride-induced testicular damage and subfertility by downregulating p53 and Bax and upregulating gonadotropins and Bcl-2 gene expression. *Journal of Reproduction and Development*, 60(2), 115–127. <https://doi.org/10.1262/jrd.2013-097>
84. Elements, T. B., Rodríguez-Álvarez, M., Paz, S., Hardisson, A., González-Weller, D., Rubio, C., & Gutiérrez, Á. J. (2011). Assessment of Toxic Metals (Al, Cd, Pb) and Trace Elements (B, Ba, Co, Cr, Cu, Fe, Mn, Mo, Li, Zn, Ni, Sr, V) in the Common Kestrel (. *Biological Trace Element Research*. <https://doi.org/10.1007/s12011-021-02974-x>
85. El-hack, M. E. A., & Alagawany, M. (2016). Review Article Nutritional , Healthical and Therapeutic Efficacy of Black Cumin (*Nigella sativa*) in Animals , Poultry and Humans. *International Journal of Pharmacology*, 12(3), 232–248. <https://doi.org/10.3923/ijp.2016.232.248>
86. Elkareem, M. A., Abd, M. A. M., Rahman, E., Khalil, N. S. A., & Amer, A. S. (2021). Antioxidant and cytoprotective effects of *Nigella sativa* L . seeds on the testis of monosodium glutamate challenged rats. *Scientific Reports*, 1–16. <https://doi.org/10.1038/s41598-021-92977-4>
87. El-kholy, A., Eraky, M., & Omar, G. (2018). *Print ISSN : 1110 - 208X Online ISSN : 2357 - 0016. January*. <https://doi.org/10.4103/bmfj.bmfj>